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The Structure of Formaldehyde-Inhibited Xanthine Oxidase Determined by 35 GHz ²H ENDOR Spectroscopy

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Abstract: The formaldehyde-inhibited Mo(V) state of xanthine oxidase (I) has been studied for four decades, yet it has not proven possible to distinguish unequivocally among the several structures proposed for this form. The uniquely large isotropic hyperfine coupling for ¹³C from CH₂O led to the intriguing suggestion of a direct Mo-C bond for the active site of I. This suggestion was supported by the recent crystal structures of glycol- and glycerol-inhibited forms of aldehyde oxidoreductase, a member of the xanthine oxidase family. ¹H and ²H ENDOR spectra of $I(C^{1,2}H_2O)$ in H_2O/D_2O buffer now have unambiguously revealed that the active-site structure of I contains a CH₂O adduct of Mo(V) in the form of a four-membered ring with S and O linking the C to Mo and have ruled out a direct Mo-C bond. Density functional theory computations are consistent with this conclusion. We interpret the large ¹³C coupling as resulting from a "transannular hyperfine interaction".

Xanthine oxidase is a molybdoenzyme that catalyzes the oxidative hydroxylation of a variety of heterocyclic and aldehyde substrates, including the physiological substrates hypoxanthine and xanthine.¹ Of the large number of EPR-active Mo(V) intermediates exhibited by this enzyme,² one of the most intriguing is the CH₂Oinhibited Mo(V) form (I) first described by Bray and co-workers.³ Its most remarkable feature, which was revealed by Howes and co-workers,^{4,5} is the presence of a carbon derived from CH₂O that EPR and ENDOR spectroscopies show to have a uniquely large ¹³C isotropic hyperfine coupling, a_{iso} (¹³C) \approx 43.0 MHz. This value contrasts with the 5-fold smaller coupling for the Mo–O–¹³C of the "2-hydroxy-6-methyl-purine (HMP) very rapid" form (a_{iso} = 7.9 MHz).⁶ Although I has been studied for 40 years, it has not proven possible to distinguish unequivocally among the viable candidates for its structure (Scheme 1).^{5,7–13}

The large isotropic ¹³C hyperfine coupling for I led Howes and co-workers to the intriguing suggestion that it contains a CHO fragment with a direct Mo–C bond (C in Scheme 1).^{4,5} This suggestion recently received support from the crystal structures of glycol- and glycerol-inhibited forms of aldehyde oxidoreductase (AOR), a member of the xanthine oxidase family.⁷ These structures exhibit Mo–C bond distances of 2.36 and 2.72 Å, respectively, the former in particular being suggestive of direct Mo–C bonding interactions. In contrast, analysis of the smaller ¹³C coupling for a substrate-derived species bound to Mo(V) of the "very rapid" state of xanthine oxidase indicated that there was no direct Mo–C bond,⁶ and this was confirmed by recent crystal structures for that intermediate.^{14,15}

Scheme 1



We here report that ¹H and ²H ENDOR spectra of $I(C^{1,2}H_2O)$ prepared¹⁶ in H₂O/D₂O buffer rule out all of the models proposed for the active-site structure of I (Scheme 1) except for model A, a four-membered cyclic adduct of CH₂O with S and O linking the C to Mo. Density functional theory (DFT) calculations are consistent with the ENDOR finding that A rather than the direct Mo–C bond of C represents the structure for the active site of I.

The reported X-band EPR spectrum of $I({}^{13}C^{1}H_{2}O)$ in $H_{2}O$ shows a doublet splitting from the ${}^{13}C$ nucleus, with each line further split into a doublet from a single proton derived from $CH_{2}O$.⁴ The 35 GHz echo-detected EPR spectrum of $I({}^{12,13}C^{1}H_{2}O)$ in $H_{2}O$ shows the ${}^{13}C$ doublet from ${}^{13}C^{1}H_{2}O$, but the ${}^{1}H$ splitting is not observed (Figure S1). 17,18 The EPR spectrum of $I({}^{12,13}C^{1}H_{2}O)$ further shows hyperfine splitting from ${}^{95,97}Mo$ (natural abundances: ${}^{95}Mo$, 15.9%; ${}^{97}Mo$, 9.6%). The g_1 and g_3 splittings are observable in both the Xand Q-band spectra; those associated with g_2 are resolved only in the Q band spectrum (Figure S1). Simulations of the EPR spectra gave principal values of the **g** tensor and values of the components of **A** for the ${}^{13}C$ of CH₂O that agree with those of Howes and coworkers: $\mathbf{g} = [1.988, 1.974, 1.948]$, $\mathbf{A}({}^{13}C) = [51.5, 40, 40]$ MHz, $a_{150} = 43.8$ MHz.

Figure 1 presents 35 GHz Davies ¹H and ^{95,97}Mo (top) and Mims ²H (bottom) ENDOR spectra collected for $I(^{12}C^{1,2}H_2O)$ in H_2O/D_2O near g_2 , the magnetic field corresponding to the maximum EPR intensity. The ¹H Davies spectra of $I(C^{1}H_2O)$ in H_2O and D_2O are essentially the same (traces **A**), both exhibiting an ¹H doublet centered at the ¹H nuclear Larmor frequency and split by $A \approx 13$ MHz. This doublet is absent in the spectrum of $I(C^{2}H_2O)$ in H_2O (trace **B**) and thus is associated with a proton, ¹H_A, derived from CH₂O. The ¹H doublet in Figure 1 rides on ^{95,97}Mo ENDOR signals and is more clearly seen in the difference spectrum **A**–**B** obtained by subtraction of the spectrum of $I(C^{2}H_2O)$ (**B**) from that of $I(C^{1}H_2O)$ (**A**).

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Figure 1. (top) Davies ¹H and ^{95,97}Mo (A, B) ENDOR spectra (35 GHz) of I(C^{1,2}H₂O) in H₂O and D₂O buffers. (bottom) Mims ²H ENDOR spectra (35 GHz) of $I(C^2H_2O)$ in H_2O (green/blue), $I(C^1H_2O)$ in H_2O (black) and D₂O (red) buffer. Horizontal bars indicate hyperfine splittings for ^{1,2}H_{A,B} nonexchangeable protons. Conditions: g = 1.97; T = 2 K. Davies: π -pulse = 80 ns, τ = 600 ns, repetition time = 50 ms, 34.84 GHz. Mims ²H: $\pi/_2$ pulse = 50 ns, τ = 800 ns, repetition time = 50 ms, 34.87 GHz.

The ${}^{1}H_{A}$ doublet is lost and is replaced by a corresponding ${}^{2}H_{A}$ signal when C^2H_2O is the substrate. ²H ENDOR spectra of $I(C^2H_2O)$ in H₂O by both Davies (not shown) and Mims (Figure 1, lower) techniques exhibit a ²H_A doublet without resolved quadrupole splitting. The hyperfine interaction, $A(^{2}H_{A})$, corresponds to a smaller value of $A({}^{1}H_{A})$ than that seen directly in the ${}^{1}H$ ENDOR spectrum of $I(C{}^{1}H_{2}O)$ in H₂O/D₂O because of a substantial isotope effect on the hyperfine couplings.^{19,20} Most importantly, the ²H Mims spectrum of I(C²H₂O) in H₂O shows doublets from both deuterons derived from the C²H₂O (Figure 1), a second doublet, denoted ${}^{2}H_{B}$, plus that from ${}^{2}H_{A}$.

To test for protons exchangeable with solvent, Mims ²H ENDOR spectra were collected for $I(C^1H_2O)$ in D_2O and $I(C^2H_2O)$ in H_2O at g_2 (Figure 1, bottom). The nearly featureless ²H spectrum of $I(C^1H_2O)$ in D_2O shows clearly that there are no exchangeable protons in the active site of I.

To confirm that the Mims ²H spectrum is not significantly distorted by suppression "holes" associated with this technique²¹ and to determine the ²H hyperfine and quadrupolar tensors, we collected a complete 2D field-frequency ²H ENDOR pattern of spectra acquired at numerous magnetic fields across the EPR envelope and simulated this pattern with inclusion of the suppression effects (Figure S2 in the Supporting Information).²² At magnetic fields near g_1 , the spectra show the ${}^{2}H_{A}$ and ${}^{2}H_{B}$ doublets centered at the ²H Larmor frequency and split by $A \approx 1.8$ and 0.4 MHz, respectively. The ²H ENDOR pattern does not change significantly as the field is moved toward g_2 and g_3 , except in intensity, indicating that the hyperfine couplings of ${}^{2}H_{A}$ and ${}^{2}H_{B}$ are largely isotropic. The 2D pattern is well-simulated by the 1:1 summation of simulations of ${}^{2}H_{A}$ and ${}^{2}H_{B}$, with $A({}^{2}H_{A}) = [1.8, 1.8, 1.9]$ MHz and $A(^{2}H_{B}) = [0.44, 0.4, 0.39]$ MHz. Neither ²H exhibits quadrupolar splitting (I = 1) because of the large ENDOR line widths. As noted above for the g_2 spectrum, the tensor obtained from simulations of the ²H_A ENDOR pattern only approximately matches that obtained by fitting a 2D ¹H Davies ENDOR pattern for ¹H_A because of an isotope effect on the hyperfine couplings.^{19,20}

The finding that both aldehydic protons of CH₂O are retained in I and are nonexchangeably hyperfine-coupled to the Mo(V) of Irules out all of the candidates for the active-site structure of I shown in Scheme 1 except the cyclic CH₂O adduct, A.

DFT calculations (SI) are consistent with experiment, yielding hyperfine tensors for ${}^{13}C$, ${}^{2}H_A$, and ${}^{2}H_B$ of CH₂O in A that all are in satisfactory agreement with the experimentally observed values: $a_{iso}(^{13}C) \approx 47.9$ MHz, $A(^{13}C) = [53.6, 45.4, 44.6]$ MHz; $A(^{2}H_{A})$ = [4.0, 3.3, 3.0] MHz; $A(^{2}H_{B})$ = [0.38, -0.67, -0.72] MHz. Most importantly, the carbon-bound CHO complex, model C, which is already ruled out by the absence of H_B, is calculated to have almost 3-fold and 5-fold smaller $^{13}\mathrm{C}$ and $^{1}\mathrm{H}_{\mathrm{A}}$ hyperfine couplings, respectively: $a_{iso}({}^{13}C) \approx 16.1$ MHz, $A({}^{13}C) = [23.2, 13.4, 11.7]$ MHz; $A(^{2}H_{A}) = [0.51, -0.43, -0.66]$ MHz.

Why is the ¹³C coupling so much larger for the carbon bound in the four-membered ring of structure A, and not directly coordinated to Mo(V), than it would be for a carbon directly bonded to the metal ion in C? We interpret the large coupling in A as resulting from a "transannular hyperfine interaction".^{23,24} The carbon is in line with a lobe of the half-occupied $Mo(d_{xy})$ orbital, and this allows overlap between $Mo(d_{xy})$ and orbitals of carbon; the large a_{iso} for 13 C corresponds to only $\sim 1.2\%$ spin density in a carbon 2s orbital. This phenonmenon was first observed for a phosphorus atom that formed part of four-membered cyclic structures of Mo(V) and V(IV) with dialkyl/aryldithiophosphinato ligands.^{23,24} A weaker hyperfine coupling was observed when the ³¹P is bound to the metal ion directly through a monodentate M-O-P linkage,²⁵ and the same is true for the Mo-O-13C of the "very rapid" intermediate.⁶ Transannular effects may have relevance to a recent proposal of a direct Fe-C bond based on a large ¹³C coupling.²⁶

Finally, regarding the use of distances in the X-ray structure to infer a direct Mo-C bond, we note that the DFT-optimized cyclic structure A has a Mo-C distance of 2.76 Å, which is the same as that found by X-ray diffraction for the glycerol-inhibited form of AOR.7

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Supporting Information Available: One EPR figure, one ENDOR figure, and DFT calculation methods. This material is available free of charge via the Internet at http://pubs.acs.org.

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